

Original Research Article

Specific Stability Indicating RP-UPLC Method for Simultaneous Estimation of Atorvastatin Calcium and Teneligliptin Hydrobromide Hydrate

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ABSTRACT

A stability-indicating RP-UPLC method was developed for the quantitative analysis of atorvastatin calcium (ATV) and teneligliptin hydrobromide hydrate (TEN) in the presence of degradants. The separation was achieved using an Acquity UPLC BEH C18 column with a mobile phase composed of Methanol, Ammonium Acetate Buffer, and Acetonitrile (60:35:5, pH 6.5, adjusted with OPA). The method was optimized with a flow rate of 1.0 mL/min, an injection volume of 2 μ L, and UV detection at 245 nm. The retention times (RT) were 2.2 min for ATV and 0.9 min for TEN. The calibration ranges were 10–30 μ g/mL for ATV and 5–15 μ g/mL for TEN, with correlation coefficients of 0.9999 and 0.9998, respectively. The method demonstrated accuracy ranging from 99.70–100.07% for ATV and 99.60–100.40% for TEN. The limits of detection (LOD) were 1.8543 μ g/mL for ATV and 0.6228 μ g/mL for TEN, while the limits of quantification (LOQ) were 1.0319 μ g/mL for ATV and 3.1271 μ g/mL for TEN. The results confirm that the developed method is robust and suitable for the analysis of a synthetic mixture in the presence of generated degradants.

Introduction

ATV is a medication that inhibits HMG-CoA reductase and is prescribed for managing hyperlipidemia. The

structure of the ATV is presented as [Figure 1 \[1\]](#). TEN is used to manage type 2 diabetes (T2D). The structure of TEN is shown in [Figure 2 \[2\]](#).

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Combination therapy can help improve lipid and glucose metabolism, as well as reduce the risk of cardiovascular disease in these patients [3]. Forced degradation, more than accelerated conditions [4], influences drug quality over time due to environmental factors [5-6]. Impurities affect both stability and pharmacology [7-12]. Analytical quality by design (AQbD) and central composite design (CCD) are instrumental in developing RP-HPLC methods, conducting stress testing, and ensuring regulatory compliance [13-15].

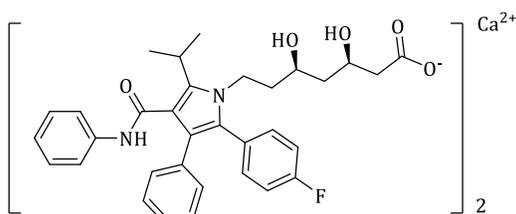


Figure 1: ATV structure

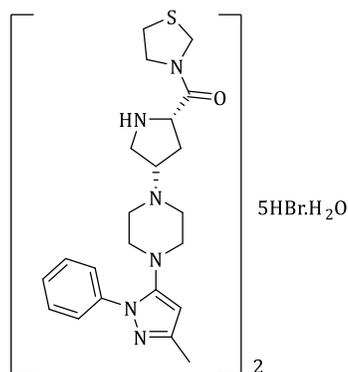


Figure 2: TEN structure

Analytical methods for drug estimation include UV spectrophotometry [16-21], RP-HPLC [22-27], stability-indicating HPLC [28-32], UPLC [33], stability-indicating UPLC [34], and LC-MS/MS [35]. One study on a stability-indicating RP-HPLC method [36] with a 17 min run time for degradation study and a 10 min RT for basic hydrolysis showed inadequate % degradation. RP-UPLC offers shorter run times, reduced solvent use, and cost savings, promoting eco-friendliness. Thus, developing and validating an isocratic RP-UPLC stability-indicating method for routine analysis is valuable.

The goal of stability and related substance studies is to demonstrate how the quality of a

drug or product changes over time due to different environmental factors [37-39]. HPLC, UV spectrophotometric methods, and LC-MS/MS are essential for quality control in pharmaceuticals, where substance stability and integrity are crucial for safety and efficacy [40-46]. The stability and pharmacological effects of active pharmaceutical ingredients (APIs) and drug products are impacted by the presence of contaminants [47-51]. AQbD and CCD help ensure adherence to regulations in the development of stability-indicating techniques, stress testing, and the RP-HPLC method [52]. RP-UPLC offers shorter run times, reduced solvent use, and cost savings, promoting eco-friendliness [53-56].

In this control structure, the central micro grid controller seeks to minimize the operation costs in the island mode and in the grid-connected mode, seeks to optimize the exchange power with the national network in order to reduce the operation costs. In this study, this type of control is used for the energy management system. In decentralized control, each micro grid is controlled by a controller. Decentralized control is a possible solution for many control and energy management

Materials and Methods

Reagents and chemicals

Standards for ATV and TEN were kindly supplied by Piramal Pharma Ltd. in Ahmedabad. HPLC-grade CH₃OH, ACN, and Milli-Q water were used, along with AR-grade ammonium acetate, HCl, NaOH, H₂O₂, and orthophosphoric acid.

Instrumentation

Chromatographic analysis was performed using a PDA detector, an Acquity UPLC-H Class, and Empower 3.6 software for data acquisition.

Chromatographic condition

UPLC was employed, and data processing was performed using Empower software. The column used for chromatographic separation was the UPLC BEH C18. The mobile phase consisted of methanol, ammonium acetate

buffer, and acetonitrile (60:35:5), with an injection volume of 2 μL and a flow rate of 1 mL/min. UV detection was carried out at 245 nm, at room temperature, with the pH adjusted to 6.5 using OPA.

Preparation of mobile phase

CH_3OH , ammonium acetate buffer and ACN were combined in a 65:30:5% v/v ratio, and then the pH was adjusted to 6.5 ± 0.05 using OPA.

Preparation of standard stock solution (200:100 $\mu\text{g/mL}$)

Approximately 20 mg of ATV and 10 mg of TEN were weighted into 100 mL of volumetric flask, and then 30 mL of diluent and sonicate were added and diluted with the diluent and mix. The resulting solution will have a concentration of 200 $\mu\text{g/mL}$ of ATV and 100 $\mu\text{g/mL}$ of TEN.

Preparation of working solution of a mixture of ATV and TEN (20:10 $\mu\text{g/mL}$)

A 50 mL volumetric flask was filled with 5 mL of ATV standard stock and 5 mL of TEN standard stock, and then diluted to the appropriate volume and mix. The resulting solution contained 10 $\mu\text{g/mL}$ of TEN and 20 $\mu\text{g/mL}$ of ATV.

Preparation of test solution

20 mg of ATV and 10 mg of TEN were weighed and diluted that resulted in concentrations of 20 $\mu\text{g/mL}$ of ATV and 10 $\mu\text{g/mL}$ of TEN.

Forced degradation studies

To evaluate the method's performance in identifying ATV, TEN, and the degradation products generated under different stress conditions, a forced degradation study was conducted.

Suitability of system

Five injections of the standard solution containing ATV and TEN were performed to verify the system's applicability. The adequacy of the system was evaluated by examining parameters such as TF, RT, and TP in the standard chromatogram.

Validation of method

By ICH (Q2R1) guidelines, analytical validation parameters were established for the examination of the suggested method.

Method validation parameters:

- Linearity
- Specificity
- LOD and LOQ
- Accuracy
- Precision
- Robustness
- Assay of synthetic mixture

Result and Discussion

Optimized condition

Extensive experimentation was conducted involving variations in mobile phase composition, column types, pH, and flow rates. The most favorable outcome was achieved using a chromatographic condition is presented in Figure 3 and Table 1.

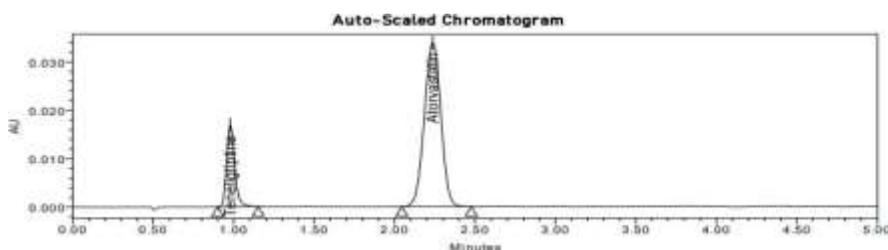


Figure 3: Chromatogram of optimized condition

Table 1: Chromatographic Condition

Sr. No.	Chromatographic parameter	Optimize condition
1	FR	1 mL/min
2	Detection λ	245 nm
3	MP composition	Methanol: Ammonium Acetate Buffer: ACN (60:35:5) (pH 6.5 Adjusted with OPA)
4	Column	Acquity UPLC BEH C18 (50 mm \times 2.1 mm, 1.7 μ m)
5	Injection volume	2 μ L
6	Column temperature	Ambient
7	RT	5 min

Linearity

The calibration curve was generated for ATV and TEN within the respective ranges of 10–30

μ g/mL and 5–15 μ g/mL, as depicted in Figures 4 and 5 as well as Table 2.

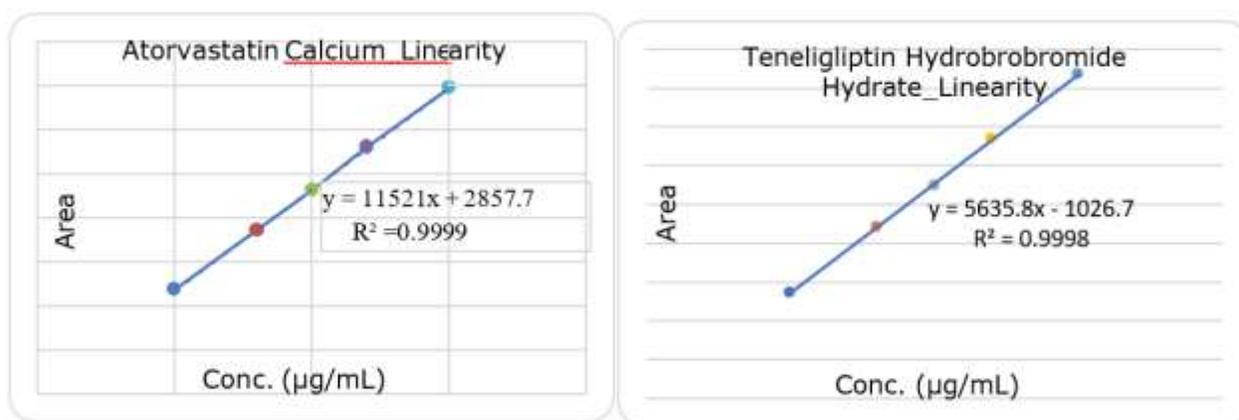
**Figure 4:** Linearity curve of ATV and TEN**Figure 5:** Linearity overlay chromatogram ATV and TEN

Table 2: Repeatability study for ATV and TEN

Drug	Y (Intercept Equation)	R ²
ATV	$y = 11521x + 2857.7$	0.9999
TEN	$y = 5635.8x - 1026.7$	0.9998

Specificity

The procedure's specificity was verified when no excipient interference was detected at the 245 nm working wavelength, and the percentage of interference for both drugs was less than 0.5%.

LOD and LOQ

The LOD and LOQ for ATV were found to be 1.0319 µg/mL and 3.1271 µg/mL, respectively, while for TEN, they were 0.6228 µg/mL and 1.8543 µg/mL.

Accuracy

The method demonstrates accuracy, with % recovery falling, as presented in [Table 3](#).

Precision

Repeatability and intermediate precision, expressed as RSD, were both below 2%, signifying the method's precision. Results are presented in [Table 3](#).

Robustness

Intentional variations in temperature, flow rate (FR), and mobile phase (MP) composition were introduced, resulting in RSD values consistently below 2%. This robustness indicates the reliability of the method and is provided in [Table 4](#).

Table 3: Accuracy, intermediate, and repeatability study for ATV and TEN

Drug Name	Level (%) (n = 3)	Interday precision RSD	Intraday precision RSD	Accuracy % drug recovery	Repeatability	
					Concentration (µg/mL) (n=6)	RSD
TEN	50	0.54	0.69	100.10	20	0.21
	100	0.35	0.36	99.87		
	150	0.22	0.28	100.04		
ATV	50	0.23	0.40	99.77	10	0.55
	100	0.14	0.18	99.87		
	150	0.13	0.18	100.04		

Table 4: Robustness study for ATV and TEN

	Flowrate (mL/min)	RSD	Wavelength (nm)	RSD	Mobile phase (MeOH: Buffer: ACN) (pH 6.5)	RSD
ATV	0.9	0.77	244	0.31	64:29:4	0.29
TEN		0.93		0.55		
ATV	1.0	0.31	245	0.35	65:30:5	0.27
TEN		0.82		0.48		
ATV	1.1	0.52	246	0.24	66:31:6	0.31
TEN		0.84		0.38		

Forced degradation study

The optimized stability-indicating approach for ATV and TEN demonstrated good separation of the drug and degradation products. The chromatogram displayed no extra peaks,

confirming specificity. Degradants were well separated, with no peak merging. Comprehensive data can be found in Table 5 and Figure 6.

Assay of synthetic mixture

The assay of a synthetic mixture containing ATV and TEN falls within an acceptable range of 98-102%. Detailed data is provided in Table 6.

Table 5: An overview of force degradation

Degradation type	% Degradation		Peak purity index	
	ATV	TEN	ATV	TEN
Acid degradation (0.1 M HCl, 8 h)	27.78	24.64	1	1
Base degradation (0.1 M NaOH, 8 h)	25.07	16.86	1	1
Oxidative degradation (3 % H ₂ O ₂ , 8 h)	18.93	25.63	1	1
Photolytic degradation (3 h)	7.23	5.15	1	1
Thermal degradation (30 min)	5.50	6.24	1	1

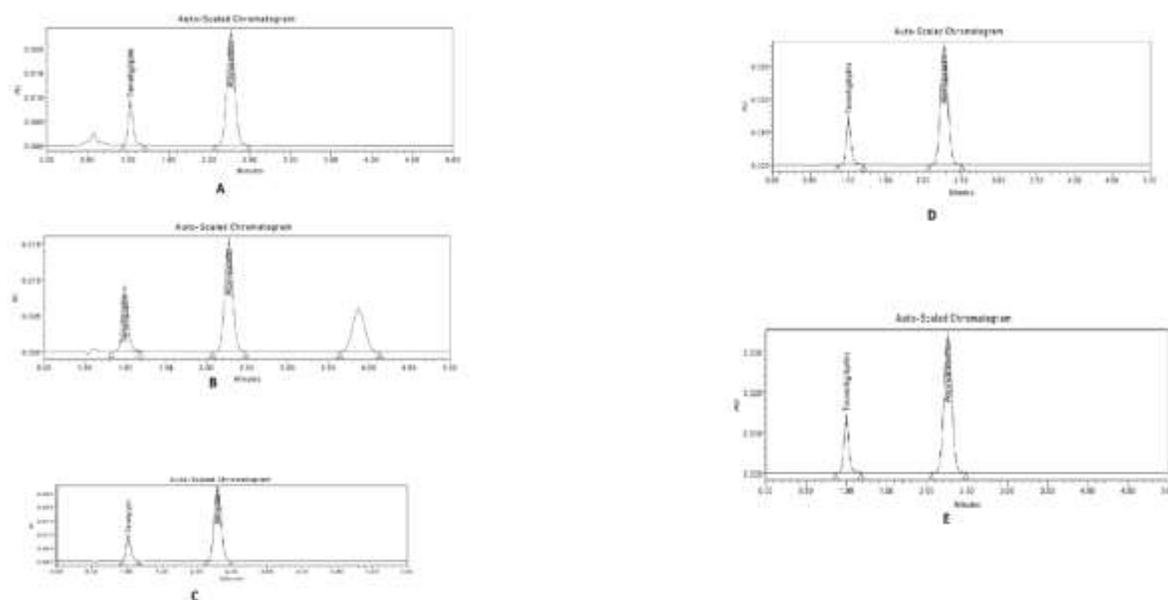


Figure 6: A) Chromatogram of acidic condition, B) chromatogram of basic condition, C) chromatogram of oxidative condition, D) chromatogram of thermal condition, and E) chromatogram of photolytic condition

Table 6: Result of assay

Conc. ($\mu\text{g/mL}$: $\mu\text{g/mL}$) ATV: TEN	Conc. found Mean \pm SD	% Drug content
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	ATV	TEN	ATV	TEN
(20:10)	19.980±0.0294	9.993±0.032	99.900	99.933

Conclusion

In a synthetic mixture, the levels of TEN and ATV were successfully measured using an RP-UPLC-PDA method that was developed and validated for stability testing. The method effectively distinguishes between the primary drugs and their degradation products. Under various stress-induced degradation conditions, ATV and TEN exhibited superior stability in thermolytic and photolytic environments compared to other stressors. Specifically, ATV demonstrated the highest stability order as thermal > photolytic > oxidation > base > acid, while TEN followed a stability order of photolytic > thermal > base > acid > oxidation. The method's specificity and selectivity were confirmed by assessing RT, resolution, and purity data for all chromatogram peaks using a PDA detector. Comprehensive validation demonstrated satisfactory results across all parameters, making this stability-indicating method suitable for pharmaceutical analysis, enabling the monitoring of drug stability and ensuring quality control.

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Conflict of Interest

The authors declare that they have no competing financial interests in this study

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Reference

- [1]. I. Pharmacopoeia, Government of India ministry of health and family welfare Ghaziabad, Published by Indian Pharmacopoeia Commission, **2010**, 1, 49-83. [Google Scholar], [Publisher]
- [2]. M. Karimi, Investigating the Microscopic Structure of Cast Iron and Its Application in Industry, *Journal of Engineering in Industrial Research*, **2023**, 2, 77-85. [Google Scholar], [Publisher]
- [3]. M.J. Daniel, Lipid management in patients with type 2 diabetes, *American health & drug benefits*, **2011**, 4, 312. [Google Scholar], [Publisher]
- [4]. M. Blessy, R.D. Patel, P.N. Prajapati, Y. Agrawal, Development of forced degradation and stability indicating studies of drugs—A review, *Journal of pharmaceutical analysis*, **2014**, 4, 159-165. [Crossref], [Google Scholar], [Publisher]
- [5]. N. Rathod, P.A.I. Prajapati, Analytical method development and validation of stability indicating method and related substance by using RP-HPLC Of drug substance, *Journal of Pharmaceutical Analysis*, 2018, 8, 367-72. [Google Scholar]
- [6]. A.J. Vyas, N.M. Visana, A.I. Patel, A.B. Patel, N.K. Patel, S.R. Shah, Analytical quality by design in stress testing or stability-indicating method, *Asian Journal of Pharmaceutical Analysis*, **2021**, 11, 170-178. [Crossref], [Google Scholar], [Publisher]
- [7]. A.B. Patel, A.H. Asnani, A.J. Vyas, N.K. Patel, A.I. Patel, D.A.N. Lumbhani, A brief review on genotoxic impurities in pharmaceuticals, *Asian Journal of Pharmaceutical Research*, **2021**, 11, 187-193. [Crossref], [Google Scholar], [Publisher]
- [8]. A.B. Patel, B.K. Jinja, A.K.J. Vyas, N.K. Patel, A.I. Patel, D.B. Sheth, S.R. Shah, A retrospective study of warning letters issued by US FDA over 2019-2021, *Asian Journal of Pharmaceutical Research*, **2022**, 12, 295-301. [Crossref], [Google Scholar], [Publisher]

- [9]. A.B. Patel, A.R. Bundheliya, A.J. Vyas, N.K. Patel, A.I. Patel, A.N. Lumbhani, A review on metal impurities in pharmaceuticals, *Asian Journal of Pharmaceutical Analysis*, **2021**, *11*, 212-222. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [10]. A.I. Patel, K.B. Prajapati, A.J. Vyas, A.B. Patel, N.K. Patel, Determination and validation of phthalate impurities in milk by UV-spectrophotometry method, *Pharma Science Monitor*, **2021**, *10*, 49-58. [[Google Scholar](#)], [[Publisher](#)]
- [11]. M. Rajput, N. Patel, U. Chotaliya, A. Patel, A. Patel, A. Vyas, Determination of genotoxic impurity by chromatographic method, *Pharma Science Monitor*, **2017**, *8*, 24-31. [[Google Scholar](#)], [[Publisher](#)]
- [12]. A.J. Vyas, S.M. Patel, A.B. Patel, A.I. Patel, N.K. Patel, S. Shah, D. Sheth, Stability testing: An Essential study for Vaccine Formulation Development, *Stress*, **2022**, *2*, R1. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [13]. A.J. Vyas, N.M. Visana, A.I. Patel, A.B. Patel, N.K. Patel, S.R. Shah, Analytical quality by design in stress testing or stability-indicating method, *Asian Journal of Pharmaceutical Analysis*, **2021**, *11*, 170-178. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [14]. A.J. Vyas, D.A. Gol, A.I. Patel, A.B. Patel, N.K. Patel, J.R. Chavda, A. Lumbhani, A. Chudasama, Implementing analytical quality by design (AQbD) approach for simultaneous estimation of tadalafil and macitentan by RP-HPLC method, *Analytical Chemistry Letters*, **2021**, *11*, 539-552. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [15]. A.I. Patel, K.B. Prajapati, S.H. Jolapara, A.J. Vyas, A.B. Patel, N.K. Patel, M.M. Pandey, RP-HPLC method for determination of gemfibrozil using central composite design (CCD), *Research Journal of Pharmacy and Technology*, **2021**, *14*, 3009-3014. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [16]. S. Naveed, Simple UV spectrophotometric assay of Atorvastatin API formulation and their comparative study, *Global Journal of Medical Research*, **2014**, *14*, 35-38. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [17]. B. Yilmaz, S. Kaban, UV and first derivative spectrophotometric methods for the estimation of atorvastatin in pharmaceutical preparations, *Journal of Advanced Pharmacy Research*, **2018**, *2*, 89-94. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [18]. S.S. Inda, S. Sharma, V. Das, B. Kumar, A. Bhandari, First derivative spectrophotometric method for the estimation of Atorvastatin calcium as bulk and in tablet dosage form, *International Journal of Pharmacy and Pharmaceutical Sciences*, **2013**, *5*, 530-533. [[Google Scholar](#)], [[Publisher](#)]
- [19]. S.A. Kshirsager, S.B. Mane, Y.S. Hanchate, A.S. Katte, K.V. Kulkarni, UV Spectrophotometric method development and validation for the determination of Teneligliptin hydrobromide hydrate in API and in pharmaceutical dosage form, *International Journal for Pharmaceutical Research Scholar*, **2018**, *7*, 19-27. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [20]. N. Yadav, A. Goyal, Method development and validation of Teneligliptin in pharmaceutical dosage form by UV spectrophotometric methods, *International Journal of Pharmaceutical Chemistry and Analysis*, **2017**, *4*, 54-58. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [21]. K. Ganesh Prabhu, Analytical method development and validation for the estimation of teneligliptin in oral solid dosage form by reverse phase chromatographic technique using UHPLC (doctoral dissertation, *RVS College of Pharmaceutical Sciences, Coimbatore*), 2017 [[Google Scholar](#)]
- [22]. S. Ertürk, E.S. Aktaş, L. Ersoy, S. Fıçıcıoğlu, An HPLC method for the determination of atorvastatin and its impurities in bulk drug and tablets, *Journal of Pharmaceutical and Biomedical Analysis*, **2003**, *33*, 1017-1023. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [23]. S.N.H. Azmi, A.K. Al-Mamari, B.S. Al-Hosni, M.R. Al-Fazari, High performance liquid chromatographic-UV method for determination of atorvastatin calcium in pharmaceutical formulations, *Journal of New Developments in Chemistry*, **2017**, *1*, 38-50. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [24]. S. Alam, S. Saleem, S. Naveed, H. Dilshad, F. Qamar, T. Alam, H. Sadia, M. Karim, A. Khan, HPLC method development and validation of atorvastatin calcium in bulk and tablet dosage form, *RADS Journal of Pharmacy and Pharmaceutical Sciences*, **2018**, *6*, 83-87. [[Google Scholar](#)]
- [25]. T.H. Atul, E.A. Rathod, K.R. Gupta, M.J.

- Umekar, HPLC and UV-spectrophotometric estimation of teneligliptin from tablet dosage form, *Asian Journal of Pharmaceutical Analysis and Medicinal Chemistry*, **2016**, *4*, 148-156. [[Google Scholar](#)], [[Publisher](#)]
- [26]. K.M. Katumala, K. Nagaraju, Analytical method development and validation for the estimation of teneligliptin and metformin Hcl by Rp-Hplc, *Indo American Journal of Pharmaceutical Sciences*, **2018**, *5*, 8385-8392. [[Crossref](#)], [[Google Scholar](#)]
- [27]. D. Gaikwad, Analytical method development and validation of teneligliptin hydrobromide in pure form by HPLC, *World Journal of Pharmaceutical Sciences*, **2017**, 37-48. [[Google Scholar](#)], [[Publisher](#)]
- [28]. Z. Zaheer, M. Farooqui, A. Mangle, A. Nikalje, Stability-indicating high performance liquid chromatographic determination of atorvastatin calcium in pharmaceutical dosage form, *African Journal of Pharmacy and Pharmacology*, **2008**, *2*, 204-210. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [29]. A. Johnson, Investigating the Effects of Environmental Applications on Decomposition of Zein Nanoparticles in Adsorbents in Industry, *Journal of Engineering in Industrial Research*, **2023**, *4*, 92-108. [[Google Scholar](#)], [[Publisher](#)]
- [30]. A. Vinit Ganorkar, R. S Jibhkate, K. Radheshyam Gupta, Development of stability indicating and robust RP-HPLC method for determination of teneligliptin, *Asian Journal of Applied Chemistry Research*, **2018**, *1*, 1-12. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [31]. M. Vaziri, S. Tabatabaee Ghomsheh, A. Azimi, M. Mirzaei, J. Khalife, Isolation and Removal of Halostonitrile from Water by Hybrid Adsorption and Nano Filtration System, *Journal of Engineering in Industrial Research*, **2021**, *3*, 114-125. [[Google Scholar](#)], [[Publisher](#)]
- [32]. T.G. Kumar, S. Vidyadhara, N.A. Narkhede, Y.S. Silpa, M.R. Lakshmi, Method development, validation, and stability studies of teneligliptin by RP-HPLC and identification of degradation products by UPLC tandem mass spectroscopy, *Journal of Analytical Science and Technology*, **2016**, *7*, 1-12. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [33]. M. Dousti, A. Firoozfar, Compacted Kerman Clay Liner: Different Permeants and Different Additives, *Journal of Engineering in Industrial Research*, **2021**, *3*, 54-68. [[Google Scholar](#)], [[Publisher](#)]
- [34]. K. Kakumani Kishore, R. Chimalakonda Kameswara, L. Maddala Vijaya, M. Khagga, A validated stability indicating RP-UPLC method for atorvastatin calcium, *American Journal of Analytical Chemistry*, **2012**, *2012*. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [35]. M. Hermann, H. Christensen, J. Reubsæet, Determination of atorvastatin and metabolites in human plasma with solid-phase extraction followed by LC-tandem MS, *Analytical and Bioanalytical Chemistry*, **2005**, *382*, 1242-1249. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [36]. U.J. Chotaliya, H.J. Dobariya, D.L. Barad, A.J. Vyas, D.A. Gol, Stability indicating RP-HPLC-DAD method for simultaneous estimation of atorvastatin calcium and teneligliptin hydrobromide hydrate in synthetic mixture, *Analytical Chemistry Letters*, **2022**, *12*, 629-638. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [37]. I.H.T. Guideline, Validation of analytical procedures: text and methodology, *Q2 (R1)*, **2005**, *1*, 05. [[Google Scholar](#)], [[Publisher](#)]
- [38]. M. Taviyad, N. Patel, U. Chotaliya, S. Singh, A. Patel, A. Patel, A. Vyas, chromatographic method development and validation for related substance, *Pharma Science Monitor*, **2017**, *8*. [[Google Scholar](#)], [[Publisher](#)]
- [39]. A.J. Vyas, C.D. Jadav, A.I. Patel, A.B. Patel, S.R. Shah, D. Sheth, S. Dholakia, Review on stability indicating assay method or forced degradation study: strategy and regulatory consideration, *Asian Journal of Pharmaceutical Analysis*, **2023**, *13*, 131-139. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [40]. A.J. Vyas, D.U. Parmar, A.B. Patel, A.I. Patel, A.V. Dudhrejya, S.R. Shah, Strategic approach for HPLC Method Development and Validation: Review, *Asian Journal of Research in Pharmaceutical Sciences*, **2024**, *14*, 71-76. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [41]. A.J. Vyas, S.A. Jha, A. Patel, A. Patel, S. Shah, D. Sheth, Review on simultaneous equation method (Vierodt's method), *Asian Journal of Pharmaceutical Analysis*, **2022**, *12*, 149-156. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [42]. A.J. Vyas, H.M. Vadile, A.I. Patel, A.B. Patel, A.V. Dudhrejya, S.R. Shah, U.J. Chotaliya, D.B. Sheth, Recent applications of UV-visible derivative spectroscopic method, *Asian Journal*

- of Pharmaceutical Analysis*, **2023**, *13*, 108-114. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [43]. A.J. Vyas, B.H. Patel, A.B. Patel, A.I. Patel, N.K. Patel, A brief review on Q-absorption ratio method in UV-spectrophotometry, *Asian Journal of Pharmaceutical Analysis*, **2022**, *12*, 281-285. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [44]. A.K.J. Vyas, S.B. Mishra, A. Patel, N. Patel, S. Shah, D. Sheth, A brief review on liquid chromatography-mass spectrometry/LCMS and its application, *Asian Journal of Pharmaceutical Analysis*, **2022**, *12*, 203-210. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [45]. A. Vaghela, A. Patel, A. Patel, A. Vyas, N. Patel, Sample preparation in bioanalysis: A review, *International Journal of Scientific & Technology Research*, **2016**, *5*, 6-10. [[Google Scholar](#)], [[Publisher](#)]
- [46]. A.J. Vyas, J.P. Godhaniya, A.I. Patel, A.B. Patel, N.K. Patel, A. Chudasama, S.R. Shah, A review on carcinogenic impurities found in marketed drugs and strategies for its determination by analytical methods, *Asian Journal of Pharmaceutical Analysis*, **2021**, *11*, 159-169. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [47]. M. Rajput, N. Patel, U. Chotaliya, A. Patel, A. Patel, A. Vyas, Determination of genotoxic impurity by chromatographic method, *Pharma Science Monitor*, **2017**, *8*, 24-31. [[Google Scholar](#)]
- [48]. A.I. Patel, A.K. Mandavia, A.J. Vyas, A.B. Patel, Nitrosamine impurity: Management of unwelcome guest in pharma market, *Asian Journal of Pharmaceutical Analysis*, **2023**, *13*, 303-308. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [49]. A.J. Vyas, S.M. Patel, A.B. Patel, A.I. Patel, N.K. Patel, S. Shah, D. Sheth, Stability testing: An essential study for Vaccine Formulation Development, *Stress*, **2022**, *2*, R1. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [50]. A.J. Vyas, N.M. Visana, A.I. Patel, A.B. Patel, N.K. Patel, S.R. Shah, Analytical quality by design in stress testing or stability-indicating method, *Asian Journal of Pharmaceutical Analysis*, **2021**, *11*, 170-178. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [51]. A.J. Vyas, D.A. Gol, A.I. Patel, A.B. Patel, N.K. Patel, J.R. Chavda, A. Lumbhani, A. Chudasama, Implementing analytical quality by design (AQbD) approach for simultaneous estimation of tadalafil and macitentan by RP-HPLC method, *Analytical Chemistry Letters*, **2021**, *11*, 539-552. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [52]. A.I. Patel, K.B. Prajapati, S.H. Jolapara, A.J. Vyas, A.B. Patel, N.K. Patel, M.M. Pandey, RP-HPLC method for determination of gemfibrozil using central composite design (CCD), *Research Journal of Pharmacy and Technology*, **2021**, *14*, 3009-3014. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [53]. A. Abdipoor, A. Taheri, A. Rangin, New magnetic graphene oxide core-shell functionalized SBA-15 dual template imprinted polymer for μ -solid phase extraction of nortriptyline and amitriptyline in mice plasma, *Separation and Purification Technology*, **2022**, *298*, 121615. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [54]. S. Akbari, A. Takhvar, E. Souri, R. Ahmadvani, A. Morsali, M.R. Khoshayand, M. Amini, A. Taheri, Comparison of testosterone extraction from human plasma using MOFs (MIL-53 (Al) and ZIF-8)-based D- μ -SPE coupled to HPLC-UV, *Chromatographia*, **2024**, 1-14. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [55]. S.H. Hassanpour, A. Alidadi, A. Doroudi, Development and validation of a reverse-phase HPLC method for determination of some water-soluble vitamins and preservatives in pharmaceutical forms, *Advanced Journal of Chemistry, Section B: Natural Products and Medical Chemistry*, **2023**, *5*, 115-129. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [56]. A. Hayat, T. Jahangir, M. Yar Khuhawar, M. Alamgir, R. Ali, A. Ali, S. Musharraf, Determination of important phenolic compounds in Pakistani brown rice varieties in controlled, germinated and fermented conditions by high performance liquid chromatography, *Progress in Chemical and Biochemical Research*, **2019**, *2*, 134-142. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]

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